www.ricerca.it Prodotti Gianni srl

VITASSAY

SARS-CoV-2+Flu A+B+RSV+Adeno Resp.

Rapid test for the simultaneous qualitative detection of nucleoprotein antigen of SARS-CoV-2, Influenza type A, Influenza type B, RSV and Adenovirus from nasopharyngeal swabs.

IUE-7715053 Ed01 October 2020





For professional in vitro diagnostic use only.

INTENDED USE

Vitassay SARS-CoV-2+ Flu A+B+RSV+Adeno Resp. is a rapid, immunochromatographic assay for the simultaneous qualitative detection of nucleoprotein antigen of SARS-CoV-2, Influenza type A, Influenza type B, RSV and Adenovirus from nasopharyngeal swabs samples from patients suspected of COVID-19 infection and/or Influenza A and/or Influenza B and/or Respiratory Syncytial Virus (RSV) and/or Adenovirus infection.

Simple, non-invasive and highly sensitivity immunoassay to make a presumptive diagnosis of SARS-CoV-2, and/or Influenza type A, influenza type B, and/or RSV and/or Adenovirus infection.

INTRODUCTION

Severe acute respiratory syndrome coronavirus 2 (SARS-Cov-2) appeared in China the first time and subsequently has spread to over 200 countries of the world with thousands of health's workers infected

Clinically, patients with SARS-Cov-2 infection tend to suffer from mile symptoms such as fever, dry cough, anosmia, fatigue, dypspnea, headache, diarrhea, and sore throat followed by vascular and systemic complications such as leukocyte infiltration of the lungs, pneumonia, severe pneumonia, severe acute respiratory diseases syndrome (ARDS), sepsis and septic shock. Recent studies in COVID-19 patients commonly manifest olfactory and gustatory dysfunction even in the absence of rhinorrhea or nasal obstruction.

The clinical presentation of respiratory infections caused by different viral pathogens can be very similar, making etiological diagnosis difficult.

Influenza virus, respiratory syncytial virus (RSV) and adenovirus are of primary importance since infections produced by them range from mild respiratory illness to fatal pneumonia, and cause considerable morbidity and excess deaths in children, elderly people, and in immunocompromised individuals throughout the world.

Influenza A and B are two types of influenza viruses that cause epidemic human disease. Uncomplicated influenza illness is characterized by the abrupt onset of constitutional and respiratory signs and symptoms (e.g. fever, myalgia, headache, malaise, nonproductive cough, sore throat, and rhinitis). Among children, otitis, nausea, and vomiting are also commonly reported with influenza illness.

RSV is a frequent cause of flu-like symptoms. It can sometimes cause lower respiratory tract illness, which can be severe, and should be considered in the differential diagnosis in such cases.

Typically adenovirus infections result in self-limiting respiratory, gastrointestinal or ocular infections, however, adenovirus can

cause severe disseminated disease in immunocompromised patients.

PRINCIPLE

Vitassay SARS-CoV-2+ Flu A+B+RSV+Adeno Resp. is a qualitative immunochromatographic assay to make a presumptive diagnosis of SARS-CoV-2, Influenza type A, Influenza type B, RSV and/or Adenovirus infection.

Strip A: The test line zone of the nitrocellulose membrane is precoated with monoclonal antibodies against SARS-CoV-2.

Strip B: The test line zone of the nitrocellulose membrane is precoated with monoclonal antibodies against Influenza type A.

Strip C: The test line zone of the nitrocellulose membrane is precoated with monoclonal antibodies against Influenza type B.

Strip D: The test line zone of the nitrocellulose membrane is precoated with monoclonal antibodies against RSV.

Strip E: The test line zone of the nitrocellulose membrane is precoated with monoclonal antibodies against Adenovirus.

During the process, the sample reacts with the antibodies against SARS-CoV-2 (strip A), Influenza A (strip B) and/or Influenza B (strip C) and/or RSV (strip D), and/or Adenovirus (strip E) forming conjugates. The mixture moves upward on the membrane by capillary action. If the sample is SARS-CoV-2 positive, antibodies present on the membrane (test line) capture the conjugate complex and a red line will be visible in the strip A. If the sample is Influenza type A positive, antibodies present on the membrane (test line) capture the conjugate complex and a red line will be visible in the strip B, if the sample is Influenza type B positive, antibodies present on the membrane (test line) capture the conjugate complex and a red line will be visible in strip C, if the sample is RSV positive, antibodies present on the membrane (test line) capture the conjugate complex and a red line will be visible in strip D and if the sample is Adenovirus positive, antibodies present on the membrane (test line) capture the conjugate complex and a red line will be visible in strip E. Although the sample is positive or negative, the mixture continues to move across the membranes and the green control line always appears (for all the strips).

The presence of these green lines (in the control zone (C)) indicates that sufficient volume is added; proper flow is obtained and serves as an internal control for the reagents.

PRECAUTIONS

- For professional in vitro use only.
- Do not use after expiration date.
- Do not use the test if its pouch is damaged.
- Clean up spills thoroughly using an appropriate disinfectant.
- Specimens should be considered as potentially hazardous and handle in the same manner as an infectious agent. A new test



must be used for each sample to avoid contaminations errors. Single use device.

- Tests should be discarded in a proper biohazard container after testing.
- Sterile swabs provided in the kits should be only used for taking the nasopharyngeal sample collection. They cannot be reuse.
- Do not touch the head of the sterile swab provided when opening their primary packaging to avoid contamination.
- Reagents contain preservatives. Avoid any contact with the skin or mucous membrane. Consult safety data sheet, available on request.
- Components provided in the kit are approved for use with the Vitassay SARS-CoV-2+ Flu A+B+RSV+Adeno Resp. Do not use any other commercial kit component.
- Follow Good Laboratory Practices, wear protective clothing, use disposal gloves, goggles and mask. Do not eat, drink or smoke in the working area.
- The presence of yellow lines in the result window (control line zone and test line zone), before using the test, is completely normal and does not imply failure of the test functionality.
- All positive results should be processed following local laws and regulations.

STORAGE AND STABILITY

Store as packaged in the sealed pouch either at refrigerated or room temperature ($2-30^{\circ}C/35.6-86^{\circ}F$).

The test is stable until the expiration date printed on the sealed pouch.

The test must remain in the sealed pouch until use.

Do not freeze.

MATERIALS

MATERIAL PROVIDED	MATERIAL REQUIRED BUT NOT PROVIDED
 10 Tests/kit Vitassay SARS-CoV-2+Flu A+B+RSV+Adeno Resp. 10 Vials with Reagent (sample diluent). 10 Swabs. Instructions for use. 	 Specimen collection container. Disposable gloves. Timer.

SPECIMEN COLLECTION

Samples should be collected in clean and dry containers.

Samples should be process as soon as possible after collection. If this is not possible, the samples can be store in the refrigerator (2- $8^{\circ}C/35.6-46.4^{\circ}F$) for 8 hours prior testing.

Samples must be brought to room temperature before testing.

Homogenize the samples as thoroughly as possible prior to preparation.

SPECIMEN PREPARATION

-Nasopharyngeal swab method:

- 1. Remove the swab from its packing.
- 2. Use the sterile swab to collect the specimen from the nostril, rotating against the nasopharyngeal wall (ensuring that swab contains cells as well as mucus).
- 3. Repeat the same procedure from the other nostril.
- 4. Process the swab as soon as possible after collecting the specimen.



PROCEDURE

Allow tests, samples and diluent to reach room temperature (15- $30^{\circ}C/59-86^{\circ}F$) prior to testing.

Do not open pouches until the performance of the assay.

-Nasopharyngeal swab method:

- 1. Open the cap of the vial for sample dilution with Reagent (figure 1).
- 2. Introduce the swab into the vial for sample dilution (figure 2) and mix the solution by rotating the swab forcefully against the side of the tube at least 1 minute. Best results are obtained when the specimen is vigorously extracted in the solution (figure 2). Extract as much liquid as possible from the swab, squeezing the sides of the tube or rotating the swab against the side of the tube as the swab is withdrawn. Discard the swab.
- 3. Close the vial with sample and diluent. Shake the vial to assure a good sample dispersion, sake during 60 seconds (figure 2).
- 4. Remove Vitassay SARS-CoV-2+ Flu A+B+RSV+Adeno Resp. from its sealed bag just before using it (figure 3).
- 5. Take the vial for sample dilution containing the diluted sample (figure 4), place it inside the multiplex tube (figure 5). Screw the cap of the multiplex tube tighly (figure 6). The bottom of the vial for sample dilution will break and the diluent+sample solution reaches the sample zone of the strips (figure 7).

Parque Tecnológico WALQA Ctra. N.330, Km.566 22197-Cuarte (Huesca, SPAIN) 6. Read the results at **10 minutes**. Do not read the test result later than 10 minutes.

If the test does not run due to solid particles (the sample is not homogenized), migration process can stop on one or more strips. In this case, tap the end of the multiplex tube on hard surface to allow migration to start again.

VIIIASSAV

4

Open the cap of the vial for sample dilution.



Introduce the swab into the vial for sample dilution and mix the solution by rotating the swab forcefully against the side of the tube. Extract as much liquid as possible form the swab. Mix 60 seconds.

 Advance Terrey
 Advance Terrey
 Advance Terrey

 Marken Terrey
 Marken Terrey
 Marken Terrey

 Mark

Vitassay SARS-CoV-2+Flu A+B+RSV+Adeno Resp.



3























Notes: The intensity of the red coloured test line in the result line region (T) will vary depending on the concentration of antigens in the specimen.

Positive results detailed in the above table should be followed up with additional confirmatory diagnostic procedures.

Single or dual simultaneous virus infections are more frequent than triple.

Invalid results: Total absence of any control coloured lines (green) indicates an invalid result, regardless of the appearance or not of the test lines (red). Wrong procedural techniques or deterioration of the reagents are mostly the main reasons for control line failure. Review the procedure and repeat the assay with a new test. If the problem persists, discontinue using the kit and contact your local distributor.

QUALITY CONTROL

Internal procedural control is included in **Vitassay SARS-CoV-2+Flu A+B+RSV+Adeno Resp.** Green line appearing in the results window is an internal control, which confirms sufficient specimen volume and correct procedural technique.

LIMITATIONS

- Vitassay SARS-CoV-2+Flu A+B+RSV+Adeno Resp. must be carried out within 2 hours of opening the sealed bag.
- The intensity of test line may vary depending on the concentration of antigens.
- The quality of Vitassay SARS-CoV-2+Flu A+B+RSV+Adeno Resp. depend on the quality of the sample; proper samples are from nasopharyngeal swabs.
- Positive results determine the presence of Influenza type A, Influenza type B, RSV and/or Adenovirus respiratory infection. A confirmed infection should only be made by a physician after all clinical and laboratory findings have been evaluated and must be based in the correlation of the results with further clinical observations.

- Positive results do not rule out co-infections with other pathogens.
- Negative results should not be considered as conclusive; it is possible that the concentration of antigen is lower than the test detection limit value. If symptoms or situation still persist, it is recommended that all negative results undergo confirmatory testing using other method and/or virus identification by cell culture and PCR.

EXPECTED VALUES

In general, most patients with COVID-19 infection only develop mild (40%) or moderate (40%) disease, 15 % develop in the severe condition that requires oxygen support, and 5% have a critical disease with complications such as respiratory distress syndrome (ARDS), sepsis and septic shock, thromboembolism, and/or multiorgan failure, including acute kidney injury and cardiac injury.

Respiratory infections caused by influenza virus type A, influenza virus type B, respiratory syncytial virus (RSV), parainfluenza virus are major causes of upper and lower respiratory tract diseases in infants and young children, causing croup, bronchiolitis, and pneumonia. Additionally, these viruses have all been identified as important causes of several lower respiratory tract diseases, with significant morbidity and mortality, in elderly and immunocompromised patients.

Sixty to ninety percent of the clinical syndrome of bronchiolitis is caused by respiratory syncytial virus (RSV) infection.

Adenoviruses are implicated in 4%-10% of cases of pneumonia, 2%-10% of cases of bronchiolitis, and 3%-9% of cases of croup.

Adenoviruses are less frequent cause of lower respiratory tract infection in children than are respiratory syncytial virus and parainfluenza virus.

PERFORMANCE CHARACTERISTICS

Analytical sensitivity (detection limit)

Detection limit value (typical value) of **Vitassay SARS-CoV-2+ Flu A+B+RSV+Adeno Resp.** (strip A: SARS-CoV-2) is 1.0 ng/mL of recombinant protein or $1 \cdot 10^3$ TCID₅₀/mL of 2019nCoV/USA-WA1/2020.

Detection limit value (typical value) of **Vitassay SARS-CoV-2+ Flu A+B+RSV+Adeno Resp.** (strip B: Influenza A) is 6.25 ng/mL of Influenza A recombinant nucleoprotein.

Detection limit value (typical value) of **Vitassay SARS-CoV-2+ Flu A+B+RSV+Adeno Resp.** (strip C: Influenza B) is 25.0 ng/mL of Influenza B recombinant nucleoprotein.

Detection limit value (typical value) of **Vitassay SARS-CoV-2+ Flu A+B+RSV+Adeno Resp.** (strip D: RSV) is 10.0 ng/mL of RSV recombinant nucleoprotein. Detection limit value (typical value) of **Vitassay SARS-CoV-2+ Flu A+B+RSV+Adeno Resp.** (strip E: Adenovirus Resp.) 1.56 ng/mL Adenovirus Hexon recombinant protein.

Clinical sensitivity and specificity

Evaluations for SARS-CoV-2+Flu A+B+RSV+Adeno Resp. (strip A_ SARS-CoV-2)

An evaluation, with 262 nasopharyngeal samples from people suspected of infection by SARS-CoV-2 virus, was performed comparing the results obtained by **Vitassay SARS-CoV-2+ Flu A+B+RSV+Adeno Resp.** (strip A) vs PCR tecnique.

Results were as follows:

		qPCR technique			
		Positive	Negative	Total	
Vitassay SARS-	Positive	26	1	27	
CoV-2+Flu A+B+RSV+Adeno	Negative	legative 2 233			
Resp (SARS-CoV-2)	Total	28	234	262	

Vitassay SARS-CoV-2+Flu A+B+RSV+Adeno Resp vs qPCR technique				
	Mean value 95% (Confidence Interval)			
Sensitivity (*)	92.9%	76.5-99.1%		
Specificity	99.6%	97.6-100.0%		
PPV	96.3%	81.0-99.9%		
NPV	99.1%	97.0-99.9%		

(*) Taking into account the recommendations for Antigendetection in the diagnosis of SARS-CoV-2 infection using rapid immunoassays (11 September 2020) from WHO, the sensitivity of the test was calculated with nasopharyngeal samples with high viral load (high viral load is expected in early symptomatic phases of the illness (with the first 5-7 days of illness) in the range of Ag-RDT test detection.

Evaluations for SARS-CoV-2+Flu A+B+RSV+Adeno Resp. (strip B and C Influenza A and Infuenza B)

Respiratory samples were used in order to evaluate the results obtained by **Vitassay SARS-CoV-2+ Flu A+B+RSV+Adeno Resp.** (strip B and C) with other immunochromatographic tests ((BinaxNOW[®] Influenza A&B (Alere).

Results were as follows:

		BinaxNOW [®] Influenza A&B		
		Positive	Negative	Total
Vitassay SARS-CoV-	Positive	5	0	5
Resp	Negative	0	6	6
(Influenza A+B)	Total	5	6	11



Vitassay SARS-CoV-2+Flu A+B+RSV+Adeno Resp vs BinaxNOW® Influenza A&B				
Sensitivity	Specificity	PPV	NPV	
>99%	>99%	>99%	>99%	

<u>Evaluations for SARS-CoV-2+Flu A+B+RSV+Adeno Resp.</u> (strip D RSV)

Respiratory samples were used in order to evaluate the results obtained by **Vitassay SARS-CoV-2+ Flu A+B+RSV+Adeno Resp.** (strip D) with other immunochromatographic tests BinaxNOW[®] RSV (Alere).

Results were as follows:

		Bin	axNOW® R	SV
		Positive	Negative	Total
Vitassay SARS-CoV-	Positive	18	0	5
2+Flu A+B+RSV+Adeno Resp (RSV)	Negative	1	10	11
	Total	19	10	29

Vitassay SARS-CoV-2+Flu A+B+RSV+Adeno Resp. vs BinaxNOW® RSV				
Sensitivity	Specificity	PPV	NPV	
95%	>99%	>99%	91%	

Evaluations for SARS-CoV-2+Flu A+B+RSV+Adeno Resp. (strip E Adenovirus Resp.)

Respiratory samples were used in order to evaluate the results obtained by **Vitassay SARS-CoV-2+ Flu A+B+RSV+Adeno Resp.** (strip E) with other immunochromatographic tests, Adenovirus Respi, (CorisBioConcept) and a immunofluorescence test (PathoDx[®]Adenovirus, Remel).

Results were as follows:

[PathoDx®Adenovirus			
		Positive	Negative	Total	
Vitaccay SARE-CoV-	Positive	20	0	20	
Vitassay SARS-CoV- 2+Flu A+B+RSV+Adeno Resp. (Adenovirus)	Negative	0	5	5	
Resp. (Adenovirus)	Total	20	5	25	
			Adenovirus Respi		
			Negative	Total	
Vitassay SARS-CoV-	Positive	20	0	20	
2+Flu A+B+RSV+Adeno Resp. (Adenovirus)	Negative	0	5	5	
	Total	20	5	25	

	Vitassay SARS-CoV-2+Flu A+B+RSV+Adeno Resp. vs PathoDx®Adenovirus Test and Adenovirus Respi Test				
Sensitivity	Specificity	PPV	NPV		
>99%	>99%	>99%	>99%		

The results showed that **Vitassay SARS-CoV-2+ Flu A+B+RSV+Adeno Resp.** has a high sensitivity and specificity to detect SARS-CoV-2, Influenza type A, Influenza type B, RSV and Adenovirus.

Hook effect

Vitassay SARS-CoV-2 does not show hook effect at:

-The concentration of SARS-CoV-2 protein tested (202500.0 $\mbox{ng/mL}).$

-The concentration of Influenza A protein tested (200000.0 $\mbox{ng/mL}).$

-The concentration of Influenza B protein tested (200000.0 $\mbox{ng/mL}).$

-The concentration of RSV protein tested (395000.0 ng/mL).

-The concentration of Adenovirus Resp. protein tested (100000.0 $\, ng/mL).$

Cross reactivity

No cross reactivity was detected against organisms that cause other respiratory infections:

Strip A: SARS-CoV-2

Adenovirus	Escherichia coli 0157	Legionella pneumophila	Salmonella enteritidis/typhi/ typhimurium/ paratyphi
Astrovirus	Entamoeba histolytica	Listeria monocytogenes	Shigella flexneri/boydii/ Sonnei/dysenteriae
Campylobacter jejuni	Giardia (CWP1, ɑ1- giardian)	Norovirus GI/Norovirus GII	Streptococcus pneumococcal
Calprotectin (human)	Helicobacter pylori	Streptococcus pneumococcal	Streptococcus pyogenes
<i>C. difficile antigen</i> GDH	Hemoglobin (human/pig Bovine)	Rotavirus	Transferrin (human)
<i>C. difficile</i> Toxin <i>A/</i> <i>C. difficile</i> Toxin B	Influenza A/ Influenza B	RSV	Yersinia O3/ Yersinia O9
Coronavirus (strain 229E, NL63, OC43)	Lactoferrin (human)		

Vitassay SARS-CoV-2+ Flu A+B+RSV+Adeno Resp. (SARS-CoV-2, strip A) showed some cross reaction with SARS and very low with MERS.

Strip B: Influenza A

Adenovirus	SARS-CoV-2 (SARS-CoV- 2)	Influenza B	RSV
Astrovirus	Coronavirus (cepas 229, NL63, OC43)	Lactoferrin (human)	Salmonella enteritidis/typhi/ typhimurium/ paratyphi
Calprotectin	Escherichia	Legionella	Shigella flexneri/boydii/



Strip C: Influenza B

Adenovirus	SARS-CoV-2 (SARS-CoV- 2)	Influenza A	RSV
Astrovirus	Coronavirus (cepas 229, NL63, OC43)	Lactoferrin (human)	Salmonella enteritidis/typhi/ typhimurium/ paratyphi
Calprotectin (human)	Escherichia coli 0157	Legionella pneumophila	Shigella flexneri/boydii/ Sonnei/dysenteriae
Campylobacter jejuni	Entamoebqa histolytica	Listeria monocytogenes	Streptococcus pneumococcal
<i>C. difficile antigen</i> GDH	Giardia (CWP1, a1- giardian)	MERS	Streptococcus pyogenes
<i>C. difficile</i> Toxin <i>A/</i> <i>C. difficile</i> Toxin B	Helicobacter pylori	Norovirus GI/Norovirus GII	Transferrina (humana)
SARS-CoV-1 (SARS)	Hemoglobin (human/pig Bovine)	Rotavirus	Yersinia O3/ Yersinia O9

Strip D: RSV

Adenovirus	SARS-CoV-2 (SARS-CoV- 2)	Influenza A/Influenza B	Salmonella enteritidis/typhi/ typhimurium/ paratyphi
Astrovirus	Coronavirus (strain 229, NL63, OC43)	Lactoferrin (human)	Shigella flexneri/boydii/ Sonnei/dysenteriae
Calprotectin (human)	Escherichia coli 0157	Legionella pneumophila	Streptococcus pneumococcal
Campylobacter jejuni	Entamoebqa histolytica	Listeria monocytogenes	Streptococcus pyogenes
<i>C. difficile antigen</i> GDH	Giardia (CWP1, a1- giardian)	MERS	Transferrin (human)
<i>C. difficile</i> Toxin <i>A/</i> <i>C. difficile</i> Toxin B	Helicobacter pylori	Norovirus GI/Norovirus GII	Yersinia O3/ Yersinia O9
SARS-CoV-1 (SARS)	Hemoglobin (human/pig Bovine)	Rotavirus	



Strip E: Adenovirus Resp.

Astrovirus	Coronavirus (strain 229, NL63, OC43)	Lactoferrin (human)	Salmonella enteritidis/typhi/ typhimurium/ paratyphi
Calprotectin (human)	Escherichia coli 0157	Legionella pneumophila	Shigella flexneri/boydii/ Sonnei/dysenteriae
Campylobacter jejuni	Entamoebqa histolytica	Listeria monocytogenes	Streptococcus pneumococcal
C. difficile antigen GDH	Giardia (CWP1, a1- giardian)	MERS	Streptococcus pyogenes
<i>C. difficile</i> Toxin <i>A/</i> <i>C. difficile</i> Toxin B	Helicobacter pylori	Norovirus GI/Norovirus GII	Transferrina (humana)
SARS-CoV-1 (SARS)	Hemoglobin (human/pig Bovine)	Rotavirus	Yersinia O3/ Yersinia O9
SARS-CoV-2 (SARS-CoV-2)	Influenza A/Influenza B	RSV	

Interference

An evaluation was performed to determine the possible interferences of **Vitassay SARS-CoV-2+ Flu A+B+RSV+Adeno Resp.** no interferences against the substances tested were detected:

Metronidazole	Loratadine	Loperamide hydrochloride (Fortasec)	Phenoxymethylp enicillin potassium
Ampicillin	Dexchlorophenira mine (Polaramine)	Heparin (Hibor)	Ambroxol hydrochloride (Mucosan)
Oseltamivir	Ebastine (Ebastel)	Almagato (Almax)	Macrogol 3350 (Movicol)
Amantadine	Acetyl Salicylic (Adiro)	Fosfamycin (Monurol)	Lysine Carbocysteinate (Pectox)
Ribavirin	Ibuprofen (Espidifen)	Aceltylcystein e (Fluimucil)	Hydroxyzine dihydrochloride
Codeine (Toseina)	Paracetamol (Dolocatil)	Dexketoprofe n trometamol (Enantyum)	Lorazepam
Benzocaine (Angileptol)	Metamizole (Nolotil)	Levofloxacin	Amoxicillin
Cloperastine (Flutox)	Prednisone	Ciprofloxacin	Mercaptopurine
<i>Carbocisteine (Iniston mucolítico)</i>	Omeprazole	Rifampicin (Rifaldin)	

REFERENCIAS/BIBLIOGRAFÍA

1. TSIVGOULIS, G; FRAGKOU, P; DELIDES, A; KAROFYLAKIS. E; DIMOPOULOU, D; SFIKAKIS, P; TSIODRAS, S. Quantitative evaluation of olfactory dysfunction in hospitalized patients with

Coronavirus [2] (COVID-19) Journal of Neurology. https://doi.org/10.1007/s00315-020-09935-9.

2. PÉREZ-GARCÍA, F; PÉREZ-TANOIRA, R; ROMANYK, J; ARROYO, T; GÓMEZ-HERRUZ, P; CUADROS-GONZÁLEZ, J. *Alltest* rapid Flow immunoassays is reliable in diagnosing SARS-CoV-2 infection from 14 days after symptom onset: A prospective single-center study. Journal of Clinicals Virology 129 (2020) 104473. https://do.org/10.1016/j.jvc.2020. 104473

3. M.T. COIRAS, J.C. AGUILAR, M.L. GARCÍA, I. CASAS, and P. PÉREZ-BREÑA, "Simultaneous Detection of Fourteen Respiratory Viruses in Clinical Specimens by Two Multiplex Reverse Transcription Nested-PCR Assays", Journal of Medical Virology, 72: 484-495, 2004

4. SCOTT A. HARPER, M.D., KEIJI FUKUDA, M.D., TIMOTHY M. UYEKI, M.D., NANCY J. COX, PhD, CAROLYN B. BRIDGES, M.D., "Prevention and Control of Influenza – Recommendation of the Advisory Committee on Immunization Practices (ACIP)", April 30, 2004 / Volume 53;1-40

5. FREYMUTH F., VABRET A., GOUARIN S., PETITJEAN J., CHARBONNEAU P., LEHOUX P., GALATEAU-SALLE F., TREMOLIERES F., CARETTE MF., MAYAUD C., MOSNIER A., BURNOUF L., "Epidemiology and diagnosis of respiratory syncytial virus in adults", Revue des Maladies Respiratoires, 2004, 21(1): 35-42

6. C. HOULIHAN, M. VALAPPIL, S. WAUGH, K. CANTLAY, DA. PRICE, "Severe adenovirus infection: an under-recognised disease with limited treatment options", JICS, Volume 13, Number 4, October 2012, pp. 337-341.

7. KATE E. TEMPLETON, SITHA A. SCHELTINGA, MATTHIAS F. C. BEERSMA, ALOYS C. M. KROES, and ERIC C. J. CLAAS, "Rapid and Sensitive Method Using Multiplex Real-Time PCR for Diagnosis of Infections by Influenza A and influenza B Viruses, Respiratory Syncytial Virus, and Parainfluenza Viruses 1, 2, 3, and 4", Journal of Clinical Microbiology, Apr. 2004, p. 1564-1569.

8. BOECK KD., "Respiratory syncytial virus bronchiolitis: clinical aspects and epidemiology", Monaldi Arch Chest Dis, 1996, Jun; 51 (3): 210-3.

9. JUNG-YUN HONG, HOAN-JONG LEE, PEDRO A. PIEDRA, EUN-HWA CHOI, KEE-HO PARK, YOUNG-YULL KOH, and WOO-SUN KIM, "lower Respiratory Tract Infections due to Adenovirus in Hospitalized Korean Children: Epidemiology, Clinical Features, and Prognosis", Clinical Infectious Diseases 2001; 32: 1423-9.

10. Antigen-detection in the diagnosis of SARS-CoV-2 infection using rapid immunoassays. Interim guidance. 11 September 2020. World Health Organization.

SYMBOLS FOR IVD COMPONENTS AND REAGENTS

IVD	in vitro diagnostic device	Ť	Keep dry
Ĩ	Consult instructions for use	X	Temperature limitation
Σ	Use by	***	Manufacturer
LOT	Batch code	Σn	Contains sufficient for <n> test</n>
DIL	Sample diluent	REF	Catalogue number



I prodotti Vitassay sono distribuiti in Italia da Prodotti Gianni srl Via Quintiliano, 30 - Milano - Italy P.Iva 08860270969 ricerca.offerte@prodottigianni.com Numero Verde: 800-832027 Fax: +39 02 5097 276

www.ricerca.it

